

Going Forward: Increasing the Accessibility of Imaging MS

Liam A. McDonnell^{1*}, Ron M.A. Heeren², Per E. André³, Markus Stoeckli⁴, Garry C. Corthals⁵,

¹ Biomolecular Mass Spectrometry Unit, Department of Parasitology, Leiden University Medical Center, Leiden, the Netherlands

² FOM Institute AMOLF, Science Park 104, Amsterdam, the Netherlands

³ Department of Pharmaceutical Biosciences, Medical Mass Spectrometry, Uppsala University, Uppsala, Sweden

⁴ Novartis Institutes of BioMedical Research, Analytical & Imaging Sciences, Basel, Switzerland

⁵ Turku Centre for Biotechnology, University of Turku & Åbo Akademi University, Turku, Finland

Running title: improving imaging MS accessibility

***Corresponding Author:**

Asst. Prof. Liam McDonnell
Biomolecular Mass Spectrometry Unit
Department of Parasitology
Leiden University Medical Center
Albinusdreef 2
2333ZA Leiden
the Netherlands
Tel: +31 (0)71 526 8744
Email: l.a.mcdonnell@lumc.nl

28

29 **Abstract**

30 The driving force behind the high and increasing popularity of imaging mass spectrometry is its
31 demonstrated potential for the determination of new diagnostic/prognostic biomarkers and its
32 ability to simultaneously trace the distributions of pharmaceuticals and their metabolites in
33 tissues without the need to develop expensive radioactively-labeled analogues. Both of these
34 applications would benefit from standardized methods, for the development of novel MS-based
35 molecular histology tests and FDA-approved MS-based assays for pharmaceutical development.
36 In addition, the broader scientific community would benefit from the increased accessibility of
37 the technique for new researchers.

38 Currently imaging MS studies are individual endeavors, utilizing the individual expertise and
39 infrastructure of a single laboratory and their immediate collaborators. A wide array of tissue preparation,
40 data acquisition and data analysis techniques have been developed but lack an international collaborative
41 structure and data sharing capabilities. Such a collaborative framework would enable methodological
42 exchange and detailed comparisons of analytical capabilities, to explore synergies between the different
43 methods and result in the development of robust standardized methods. Here we describe a new European
44 imaging MS network that will explicitly compare and contrast existing methods to provide best practice
45 guidelines for the entire healthcare research community.

46

47

48

49

50

51

52 **Introduction**

53 The ability of modern proteomic techniques to identify and quantify the levels of thousands of proteins
54 from a tissue or biofluid sample has transformed medical and biological research. In addition to global
55 protein profiling, methods have been developed that target the study of protein isoforms, specific
56 pathological entities or subcellular components. It has been increasingly recognized how the rich
57 molecular information provided by this array of approaches offers new possibilities in the clinical field.
58 This ranges from new insights into the molecular changes associated with pathogenesis, the identification
59 of new therapeutic targets, to improved diagnosis and prognosis through the determination of biomarker
60 proteins and protein profiles associated with a disease and its progression, respectively.

61 Many international clinical institutions have initiated research programs for the identification of biomarker
62 protein and protein profiles, mostly in easily accessible body fluids such as plasma or serum [1].

63 Difficulties associated with dilution of the biomarkers in these fluids and the intrinsic variability of the
64 protein content of body fluids [2-4] have led to an alternative strategy: searching for biomarkers directly in
65 the affected tissue. Recent notable examples include a combination of tissue microdissection, protein
66 extraction, and extensive peptide/protein separation to quantitatively investigate the changes in protein
67 content associated with the different stages of pancreatic intraepithelial neoplasia [5] and multiple
68 sclerosis [6].

69 Mass spectrometry (MS) based methods can be directly applied to tissue [7, 8]. Imaging MS can
70 be considered spatially resolved direct tissue analysis, and uses the same tools to simultaneously analyze
71 the distribution of hundreds of biomolecules in tissue [9-11]. Rapid progress in the field now allows such
72 multiplex imaging studies to be performed with high sensitivity and selectivity. Using complementary
73 sample preparation strategies imaging MS can be used to analyze peptides, proteins, metabolites and
74 pharmaceuticals, without labeling and without prior knowledge [12-15]. When combined with a
75 histological analysis of clinical tissues imaging MS can be used to identify and visualize differentially

76 expressed biomarkers [11, 16, 17], *e.g.* Walch and workers have demonstrated that imaging MS could
77 identify features that differentiated between six different tumors [18].

78 Several studies have already shown how imaging MS can be used to chart biomolecular variation
79 in clinical tissue samples: many candidate peptide/protein/metabolite biomarkers have been identified,
80 including markers that discriminate between clinically challenging pathologies [19, 20]. This spatio-
81 chemical direct tissue analysis has the potential to bring modern biomolecular techniques into the clinic by
82 providing a biomolecular screening that complements histopathological analysis.

83 The impact of imaging MS for clinical and pharmaceutical research is reflected in its increasing
84 use throughout international clinical and pharmacological institutions and the availability of imaging mass
85 spectrometers, tissue preparation stations and data analysis solutions from leading instrument
86 manufacturers. A formidable array of capabilities has now been developed: high spatial resolution [14, 21-
87 24], high mass resolution [25, 26], multiple molecular classes [12, 26, 27], high sensitivity and specificity,
88 in-tissue identification[28-31] and quantitation [32, 33], high throughput analyses [34], and integration
89 with established pathological [35, 36] and ‘omics methodologies [37, 38]. Researchers utilizing imaging
90 MS in healthcare research would benefit enormously from a detailed comparison of many of the above
91 tools; the identification of synergies could provide new diagnostic tests for a range of pathologies as well
92 as new tools for pharmaceutical development. Importantly, such a comparison would also provide a
93 testing ground for the applicability of the technology for a range of pathologies and help develop
94 standardized methodologies for its wider implementation and improve its accessibility.

95

96 *Current state of knowledge*

97 Imaging mass spectrometry has enabled the levels and distributions of panels of biomolecules to be
98 simultaneously measured in tissues. The goal has been to exploit the intrinsic capabilities of mass
99 spectrometry for pathohistological and pharmaceutical analysis, specifically:-

- 100 • Analysis of multiple molecular classes.

- 101 • Parallel analysis of panels of biomolecules containing 100's of distinct species. For example the
102 application of an automated peak detection routine to the imaging MS datasets of myxofibrosarcoma
103 tissue samples identified 358 discrete peptide and protein peaks [39].
- 104 • Ability to distinguish between isoforms by exploiting the difference in their mass.
- 105 • Improved relative quantitation.
- 106 • Label free analysis – thus prior knowledge of the biomolecular content is not required.

107

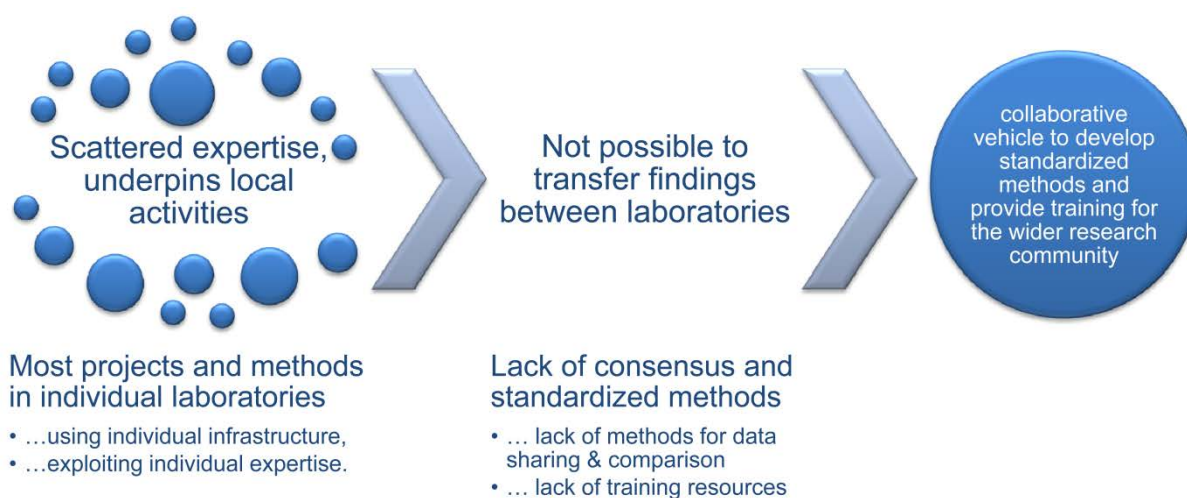
108 An array of methods has been developed to achieve these goals:-

- 109 • Sample preparation strategies to enable the analysis of peptides, proteins, lipids, pharmaceuticals and
110 metabolites [40].
- 111 • Mass analysis methods optimized for sensitivity, spatial resolution [14, 22, 24], mass resolution [25,
112 26], high mass molecules [41], or combined with an additional ion-separation dimension [42, 43].
- 113 The review by Heeren and co-workers contained in this special issue contains a detailed overview of
114 the technological developments that have been undertaken in one of Europe's principal MS
115 technology development laboratories [44].
- 116 • A host of data reduction and data analysis capabilities (see review by Jones *et al.* in this special issue
117 [45]).
- 118 • Combined imaging MS – histological/histochemical analysis of tissues for the identification of
119 candidate biomarkers [8, 18, 35, 36, 46-51].
- 120 • Imaging MS based molecular histology for revealing molecular changes that occur prior-to/without
121 morphological change [19, 20, 39, 52-55].
- 122 • Correlation with patient outcome for the identification of prognostic biomarkers [47, 56].
- 123 • Correlation with patient response-to-therapy, thereby potentially aiding personalized medicine [57].

124

125 The reviews contained in this special issue demonstrate the rapid progress being made in the field and
 126 provide ample evidence that imaging MS can complement established histological and histochemical
 127 methods and aid patient or disease stratification. Despite the increasing use of imaging MS and its
 128 widespread commercial availability the majority of studies are independent research projects, utilizing
 129 individual infrastructure and based on the individual expertise of the research group and their
 130 collaborators. The scattered nature of the expertise, the lack of methods for data sharing (beginning to be
 131 addressed by the *imzML* data standard [58]) and suitable training has severely limited the ease of
 132 knowledge transfer between laboratories, Figure 1. Currently, robust imaging MS experiments across
 133 multiple laboratories are conspicuous by their absence.

134



135

136 **Figure 1.** A collaborative vehicle is necessary to overcome the current individualist nature of
 137 imaging MS experiments and lack of data sharing/comparison tools.

138

139 *The need for improved accessibility*

140 Modern mass spectrometers provide exquisite mass analysis capabilities: high sensitivity, high
 141 resolution and high mass accuracy enable many biomolecules to be characterized directly from

142 tissue. The review by Goodwin clearly communicates the importance of sample preparation to
143 the success of an imaging MS experiment and the need to control as many variables as possible
144 (“small mistakes can lead to big consequences” [59]). Many reviews have focused on sample
145 preparation [13, 40], which reflects its critical role in determining the quality of imaging MS
146 data. In 2007 a discussion began concerning the merits of a database containing sample
147 preparation protocols to enable new researchers working in the healthcare community to exploit
148 the technology. The problem of a lack of standardization and training in sample preparation was
149 compellingly confirmed during the first Nordic Signals imaging MS training course (organized
150 by Corthals, McDonnell and Heeren and held at the FOM Institute AMOLF in March 2009):
151 despite most participants using the same commercial matrix deposition device its practical usage
152 differed widely (and in most cases contrary to the manufacturer’s recommendations). With
153 suitable training all participants were able to generate near identical, high quality imaging MS
154 datasets describing the molecular content of the tissue.

155 A similar situation was evident in the subsequent data analysis training course (organized
156 by Corthals and McDonnell and held at Turku Centre for Biotechnology in December 2009). The
157 majority of participants were restricted to the data analysis tools included in the software
158 provided with their commercial instruments or the freely available Biomap software, an
159 independent package developed for imaging MS by Markus Stoeckli (and which has been an
160 essential element in the growth of the technique). Nevertheless the single biggest impediment to
161 effective data analysis was a lack of training to fully utilize the statistical methods that were
162 available. Following the training course the participants were able to generate robust classifiers
163 and perform molecular histology much more effectively.

164 The restriction to data analysis packages supplied with the commercial mass
165 spectrometers or Biomap means that most researchers have not been able to exploit the improved

166 data analysis capabilities reported by data analysis specialists [45]. A lack of a data format
167 standard is one, significant, reason most newly reported capabilities are not utilized as much as
168 they may otherwise be; an open-source imaging MS data analysis platform that incorporates the
169 latest algorithms would certainly help but would only be part of the answer: the researchers must
170 understand the data analysis algorithms in order to use them correctly. The lengthy introduction
171 to imaging MS data analysis [45] contained in this review is meant to begin to address this need.

172 Only with sufficient training and co-operation can the full potential of imaging MS be
173 utilized to test the capabilities of these highly cross-disciplinary tools against an array of diseases
174 of present day concern, both in terms of improved diagnosis and pharmacological development.
175 Central to this purpose will be the dissemination of the complementary techniques and expertise
176 and their sustained interaction. Interaction between imaging MS researchers is crucial for
177 devising best practice guidelines and web-based experimental resources; the involvement of
178 healthcare researchers is essential in order to ensure the imaging MS efforts target real needs in
179 healthcare research, e.g. differentiation patient subgroups, and pharmaceutical development, e.g.
180 more cost-effective methods for differentiation of lead compounds.

181

182 *Increasing Accessibility of Imaging MS*

183 A large European network was recently announced, COST Action BM1104, entitled Mass Spectrometry
184 Imaging: New Tools for Healthcare Research, the sole aim of which is to establish imaging mass
185 spectrometry and related translational technologies in clinical and pharmaceutical research. Best
186 practice guidelines for data acquisition will be created for describing tissues by their molecular
187 content and distribution, which will then be exploited to develop new molecular histological
188 signatures for improved disease diagnosis as well as new methods for quantitative imaging of
189 lead formulations for pharmaceutical development. Importantly the network is also supported by

190 the European Proteomics Association, which has made Imaging MS one of four special initiatives
191 to be promoted. The central idea behind the COST Action is information exchange and training;
192 for data acquisition, data analysis, and their application and to provide this knowledge as public
193 resources, Figure 2. The generous hosting of the Action on the website www.maldi-msi.org, the
194 de-facto online center for imaging MS news and resources, will further increase the networks
195 visibility and impact.



196
197 **Figure 2.** The four elements of COST Action BM1104, designed to improve the accessibility and
198 interlab reproducibility of imaging MS.

199
200 *Data Acquisition – Best Practice Guidelines*
201 Imaging MS requires the localized extraction of the molecules of interest followed by spatially
202 correlated mass analysis. The sample preparation and mass analysis methods are critical factors
203 that determine which molecules are measured, and the sensitivity and resolution at which they
204 can be detected. Imaging MS and healthcare researchers will visit each other’s laboratories to test
205 the performance of the imaging MS methods (sample preparation, mass analysis) that have been

206 developed in each laboratory. The explicit inclusion of multiple pathologies and multiple
207 practitioners provides the capacity and redundancy to begin devising best practice guidelines for
208 multiple molecular classes and tissues.

209

210 *Data Analysis*

211 Histology-defined analysis can be used for the identification of biomarkers specific to
212 pathological entities, and histology-independent analyses examine and classify the tissue solely
213 on the basis of their MS signatures [45]. Both of these approaches have the potential to generate
214 new diagnostic tools and many data analysis techniques have been developed. However as most
215 imaging MS experiments have been performed using commercial instruments using proprietary
216 data formats many clinical users have been ‘locked’ into single data analysis packages and have
217 not been able to exploit the new data analysis capabilities.

218 A new imaging data standard, *imzML*, was developed within the 6th framework program
219 Computis [58]. Substantial support from instrument vendors has led to *imzML* being implemented
220 as an export option on most commercially available instruments. The different data analysis
221 capabilities developed in the partner laboratories will be made *imzML* compliant to enable
222 widespread data sharing and an explicit comparison of the different imaging data analysis tools,
223 Figure 3. Comparing the performance of the data analysis tools for a variety of pathologies will
224 establish standardized tools and *context-dependent* best-practice guidelines for analyzing such
225 rich datasets.

226

227

228

229



230

231 **Figure 3.** The imaging MS data standard, *imzML*, will be adopted by European COST Action
232 BM1104 to enable data sharing and a comparison of data analysis algorithms, both essential
233 elements for the development of robust imaging MS data analysis strategies and improving the
234 accessibility of the technique to new researchers.

235

236 The improvements in sensitivity, quantitation, data analysis and data sharing provided by this
237 research network could benefit many application areas. A database of sample preparation
238 protocols and data analysis strategies together with tools for data sharing, and demonstrated on a
239 selection of pharmacological and pathological applications, will be made publicly available to
240 enable further investigations in healthcare research.

241

242 The explicit production of databases detailing effective experimental protocols for imaging MS
243 analysis of tissue will provide a valuable resource to the mass spectrometry community that is
244 currently lacking, which is crucial both for the wide exploitation of this up-and-coming technique
245 but also for the development of robust inter-laboratory validated imaging-MS based assays.

246

247 *Imaging MS Standardization and Method Validation*

248 The primary bioanalytical goal of the COST Action is improved imaging MS analysis through an
249 extensive comparison of the imaging MS methods that have been developed in European
250 laboratories. No less important are the logical consequences of this goal:-

- 251 i) High performance imaging MS throughout member laboratories: the research network will
252 help maintain optimum performance of the imaging MS infrastructure. Instead of the
253 somewhat overly-reductivist question of ‘which is the best instrument?’, and which ignores
254 the high expense of mass spectrometers, the training, best practice guidelines and open
255 availability of high performance data analysis algorithms will mean that researchers can use
256 their current infrastructure to its maximum capability. This aspect can be considered the first
257 imaging MS multicentre standardization initiative, similar to the recently reported protein
258 identification and quantitation inter-laboratory comparisons [60, 61], and will build on the
259 substantial expertise of the Spanish Proteored organization [62].
- 260 ii) Inter-laboratory validation of imaging MS based assays. The development of clinical assays
261 based on imaging MS needs to explicitly assess if the specific biomarkers or biomarker
262 profiles can be detected in multiple laboratories (equipped with similar technologies) and
263 using multiple tissue banks. The final aspect is crucial because of differences in tissue
264 collection/storage may affect the MS profiles recorded by MALDI imaging MS. The COST
265 research network provides the research capacity to begin to address this challenge:

- 266 a. Researchers from laboratory *A*, using tissues from tissue bank *A*, can visit partner
267 laboratories to assess if the biomarkers determined by laboratory *A* are also
268 effective when the imaging MS experiments are performed at partner laboratories
269 *B*, *C* and *D*.
- 270 b. Researchers from laboratory *B*, *C* and *D*, using tissues from their own tissue banks
271 can visit partner laboratory *A* to determine if the biomarkers determined in
272 laboratory *A* are effective using independent tissue collections.

273 The separation of tissue source and imaging MS technique, two potentially significant
274 sources of variation, is essential for the establishment of robust diagnostic tests based on
275 imaging MS. Without standardization imaging MS will be limited to a biomarker discovery
276 role in which the final validation, and any subsequent clinical assay, is performed using an
277 alternative technique. Immunohistochemistry can, and will, be used for individual proteins if
278 suitable antibodies are available. However mass spectrometry is much more sensitive to
279 protein isoforms and so great care will need to be taken to ensure the antibody assays target
280 the isoforms of interest. For example a recent report concerning prostate specific antigen
281 [63] has highlighted that though multiple forms of PSA could be identified using MS
282 methods, and which were demonstrated to be enzymatically active and could differentiate
283 patient groups, an established antibody-based clinical method did not detect many of the
284 isoforms and underestimated the PSA heterogeneity. Similarly whereas the change in mass
285 associated with neuropeptide isoforms makes them readily distinguishable by imaging MS
286 [64], high specificity antibodies for distinct isoforms may not be available. For metabolites
287 and lipids, it is often the case that imaging MS provides the only method by which the
288 distributions of specific species may be obtained [20, 38, 65]. In short the creation of

289 standardized imaging MS methods, that demonstrate high reproducibility in multi-centre
290 experiments, has enormous potential to offer new molecular histology capabilities.

291

292 *Target groups*

293 There are several target groups and end users that will benefit from this Action:-

294 i) Healthcare researchers will benefit from improved and standardized methods for molecular
295 histological analysis of pathological tissues. Annual training courses will provide the academic
296 background and hands-on training to enable its rapid implementation. This will provide new
297 methods for biomarker discovery and new diagnostic tools to complement established
298 histopathological analysis.

299 ii) Pharmacological researchers and industry will benefit from improved and standardized
300 methods for analyzing pharmaceuticals and the metabolites in tissues. This will provide a more
301 rapid and cost effective method for testing the efficacy of lead compounds during drug
302 development.

303 iii) Mass spectrometry researchers and industry will benefit from an array of tools that have been
304 designed for analyzing the large datasets generated by imaging mass spectrometry and data
305 sharing platforms that enable efficient interaction with external collaborators.

306 iv) Imaging mass spectrometry researchers will benefit from the standardized protocols, training
307 and tools generated by the Action. Improved sensitivity, integration of data from multiple
308 molecular classes, efficient data analysis and data sharing methods, will provide the enabling
309 technologies for the widespread application of these tools to a diverse array of pathological
310 questions.

311 v) Standards Bodies: the data sharing methodologies that will be developed will provide new data
312 standards and data analysis standards for imaging MS.

313 vi) Industry: databases of effective protocols covering all aspects of the experiment, standardized
314 data analysis tools and targeted training courses will enable the tools developed with the Action
315 to be tested and implemented more rapidly and cost effectively than is currently possible. These
316 tools can be directly applied to healthcare and pharmacological industries, and could be adapted
317 to fields as diverse as food development and synthetic materials.

318 The above list of benefits for specific groups of researchers underestimates the potential impact
319 of the network. The network will be greater than the sum of its parts (groups of researchers)
320 because of the high degree of complementary expertise; creating a community of researchers that
321 actively share, discuss and generate new ideas will propel imaging MS into the healthcare field,
322 as well as enrich the experiences and careers of the participants.

323

324 **Concluding remarks**

325 Imaging MS is now beginning to deliver its potential in the clinical and pharmaceutical fields.
326 Multiple examples of semi-quantitative and quantitative imaging of drugs and tracing agents
327 (PET, MRI) have been reported, and a number of histology-defined clinical imaging MS studies
328 have reported biomarkers for improved diagnosis and prognosis. More slowly, imaging MS-
329 based molecular histology tools are emerging to define tissues on the basis of their MS profiles,
330 thereby potentially complementing established histological and histochemical methods.
331 Imaging MS is approaching a crossroad. The field may continue on its current path, in which
332 most experiments utilize individual infrastructure, tissue resources and expertise – more
333 candidate biomarkers will be reported, some proteins will be independently validated by applying
334 immunohistochemistry to tissue-microarrays and metabolites/lipids/pharma will be validated
335 using LC/CE-MS analysis of tissue extracts. The recent commercial availability of liquid
336 extraction surface analysis provides a more routine method for localized sampling of tissues [66].

337 While such individual investigations will undoubtedly uncover a veritable trove of results the
338 individual nature of the experiments will mean imaging MS is limited to a discovery role. Only
339 by developing standardized and robust methods that can be *routinely implemented in a clinic*,
340 with sufficient throughput to analyze tissue series (biomarker discovery) and speed (patient
341 diagnosis) can the full potential of imaging MS begin to be harnessed. An essential element will
342 be to ascertain which biomarkers, and biomarker profiles, can be internationalized. The other
343 path in the cross-roads is to accept that such an investigation is better achieved as a community.
344 The imaging MS community of researchers brought together within COST Action BM1104, in
345 which each participant brings a willingness to compare and contrast methods that have been
346 painstakingly developed in their laboratory, is hoped to act as a springboard for imaging MS to
347 make the transition from an up-and-coming technique with high potential, to an established
348 biomolecular histological tool with high utility in both the healthcare and pharmacological fields.

349

350 **Acknowledgements**

351 The research network is funded through COST Action BM1104, entitled Mass Spectrometry
352 Imaging: New Tools for Healthcare Research, and is gratefully acknowledged by all authors.

353

354 **Figure Captions**

355 **Figure 1.** A collaborative vehicle is necessary to overcome the current individualist nature of
356 imaging MS experiments and lack of data sharing/comparison tools.

357 **Figure 2.** The four tongs of COST Action BM1104, designed to improve the accessibility and
358 interlab reproducibility of imaging MS.

359 **Figure 3.** A common data standard, *imzML*, enables data sharing and international comparison of
360 data analysis algorithms, essential elements for the development of optimum and robust imaging
361 MS data analysis strategies.

362

363 **References**

- 364 [1] Wright ME, Han DK, Aebersold R. Mass Spectrometry-based Expression Profiling of Clinical Prostate
365 Cancer. *Mol Cell Proteomics*. 2005;4:545-4.
- 366 [2] Gerszten RE, Accurso F, Bernard GR, Caprioli RM, Klee EW, Klee GG, et al. Challenges in translating
367 plasma proteomics from bench to bedside: update from the NHLBI Clinical Proteomics Programs. *Am J*
368 *Physiol-Lung C*. 2008;295:L16-L22.
- 369 [3] Hortin GL, Sviridov D. The dynamic range problem in the analysis of the plasma proteome. *J*
370 *Proteomics*. 2009;73:629-36.
- 371 [4] Pieragostino D, Petrucci F, Boccio PD, Mantini D, Lugaresi A, Tiberio S, et al. Pre-analytical factors in
372 clinical proteomics investigations: Impact of ex vivo protein modifications for multiple sclerosis biomarker
373 discovery. *J Proteomics*. 2009;73:579-92.
- 374 [5] Sitek B, Sipos B, Alkatout I, Poschmann G, Stephan C, Schulenburg T, et al. Analysis of the Pancreatic
375 Tumor Progression by a Quantitative Proteomic Approach and Immunohistochemical Validation. *J*
376 *Proteom Res*. 2009;8:1647-56.
- 377 [6] Han MH, Hwang S-I, Roy DB, Lundgren DH, Price JV, Ousman SS, et al. Proteomic analysis of active
378 multiple sclerosis lesions reveals therapeutic targets. *Nature* 2007;451:1076-81.
- 379 [7] Schwartz SA, Weil RJ, Thompson RC, Shyr Y, Moore JH, Toms SA, et al. Proteomic-Based Prognosis of
380 Brain Tumor Patients Using Direct-Tissue Matrix-Assisted Laser Desorption Ionization Mass
381 Spectrometry. *Cancer Res*. 2005;65:7674-81.
- 382 [8] Yanagisawa K, Shyr Y, Xu BJ, Massion PP, Larsen PH, White BC, et al. Proteomic Patterns of Tumour
383 Subsets in Non-Small-Cell Lung Cancer. *Lancet*. 2003;362:433-9.
- 384 [9] Cornett DS, Reyzer ML, Chaurand P, Caprioli RM. MALDI Imaging Mass Spectrometry: Molecular
385 Snapshots of Biochemical Systems. *Nat Methods*. 2007;4:828-33.
- 386 [10] McDonnell LA, Heeren RMA. Imaging Mass Spectrometry. *Mass Spectrom Rev* 2007;26:606-43.
- 387 [11] Schwamborn K, Caprioli RM. Molecular imaging by mass spectrometry - looking beyond classical
388 histology. *Nat Rev Cancer*. 2010;10:639-46.
- 389 [12] Eijkel GB, Kùkrer Kaletaş B, van der Wiel IM, Kros M, Luider TM, Heeren RMA. Correlating MALDI and
390 SIMS Imaging Mass Spectrometric Datasets of Biological Tissue Surfaces. *Surf Interface Anal*.
391 2009;41:675-85.
- 392 [13] Kaletas BK, van der Wiel IM, Stauber J, Dekker LJ, Guzel C, Kros JM, et al. Sample preparation issues
393 for tissue imaging by imaging MS. *Proteomics*. 2009;9:2622-33.
- 394 [14] Ròmpp A, Guenther S, Schober Y, Schultz O, Takats Z, Kummer W, et al. Histology by Mass
395 Spectrometry: Label-Free Tissue Characterization Obtained from High-Accuracy Bioanalytical Imaging.
396 *Angew Chem Int Ed*. 2010;49:3834-8.
- 397 [15] Seeley EH, Oppenheimer SR, Mi D, Chaurand P, Caprioli RM. Enhancement of protein sensitivity for
398 MALDI imaging mass spectrometry after chemical treatment of tissue sections. *J Am Soc Mass Spectrom*.
399 2008;19:1069-77.

400 [16] McDonnell LA, Willems SM, Corthals GL, van Remoortere A, van Zeijl RJM, Deelder AM. Imaging
401 Mass Spectrometry in Cancer Research: Past Experiences and Future Possibilities. *J Proteomics*.
402 2010;73:1921-44.

403 [17] Rauser S, Deininger S-O, Suckau D, Höfler H, Walch A. Approaching MALDI Molecular Imaging for
404 Clinical Proteomic Research: Current State and Fields of Application. *Expert Rev Proteomic*. 2010;7:927-
405 41.

406 [18] Meding S, Nitsche U, Balluff B, Elsner M, Rauser S, Schöne C, et al. Tumor Classification of Six
407 Common Cancer Types Based on Proteomic Profiling by MALDI Imaging. *J Proteome Res*. 2012;11:1996-
408 2003.

409 [19] Lazova R, Seeley EH, Keenan M, Gueorguieva R, Caprioli RM. Imaging mass spectrometry-a new and
410 promising method to differentiate spitz nevi from spitzoid malignant melanomas. *Am J Dermatopathol*.
411 2012;34:82-90.

412 [20] Willems SM, van Remoortere A, van Zeijl R, Deelder AM, McDonnell LA, Hogendoorn PCW. Imaging
413 Mass Spectrometry of Myxoid Sarcomas Identifies Proteins and Lipids Specific to Tumor Type and Grade,
414 and Reveals Biochemical Intratumor Heterogeneity. *J Pathol*. 2010;222:400-9.

415 [21] Broersen A, van Liere R, Altalear AFM, Heeren RMA, McDonnell LA. Automated, Feature-Based
416 Image Alignment for High-Resolution Imaging Mass Spectrometry of Large Biological Samples. *J Am Soc*
417 *Mass Spectrom*. 2008;19:823-32.

418 [22] Lagarrigue M, Becker M, Lavigne R, Deininger S-O, Walch A, Aubry F, et al. Revisiting Rat
419 Spermatogenesis with MALDI Imaging at 20 μ m Resolution. *Mol Cell Proteomics*. 2011;10:M110.005991.

420 [23] Luxembourg SL, McDonnell LA, Mize TH, Heeren RMA. Infrared Mass Spectrometric Imaging below
421 the Diffraction Limit. *J Proteome Res*. 2005;4:671-3.

422 [24] Luxembourg SL, Mize TH, McDonnell LA, Heeren RMA. High-Spatial Resolution Mass Spectrometric
423 Imaging of Peptide and Protein Distributions on a Surface. *Anal Chem*. 2004;76:5339-44.

424 [25] Cornett DS, Frappier SL, Caprioli RM. MALDI-FTICR Imaging Mass Spectrometry of Drugs and
425 Metabolites in Tissue. *Anal Chem*. 2008;80:5648-53.

426 [26] Taban IM, Altalear AFM, Fuchser J, van der Burgt YEM, McDonnell LA, Baykut G, et al. Imaging of
427 Peptides in the Rat Brain Using MALDI-FTICR Mass Spectrometry. *J Am Soc Mass Spectrom*. 2006;18:145-
428 51.

429 [27] Römpf A, Guenther S, Takats Z, Spengler B. Mass spectrometry imaging with high resolution in mass
430 and space (HR2 MSI) for reliable investigation of drug compound distributions on the cellular level. *Anal*
431 *Bioanal Chem*. 2011;401:65-73.

432 [28] Franck J, Arafah K, Barnes A, Wisztorski M, Salzet M, Fournier I. Improving Tissue Preparation for
433 Matrix-Assisted Laser Desorption Ionization Mass Spectrometry Imaging. Part 1: Using Microspotting.
434 *Anal Chem*. 2009;81:8193-202.

435 [29] Groseclose MR, Andersson M, Hardesty WM, Caprioli RM. Identification of proteins directly from
436 tissue: in situ tryptic digestions coupled with imaging mass spectrometry. *J Mass Spectrom*. 2007;42:254-
437 62.

438 [30] Groseclose MR, Massion PP, Chaurand P, Caprioli RM. High-Throughput Proteomic Analysis of
439 Formalin-Fixed Paraffin-Embedded Tissue Microarrays Using MALDI Imaging Mass Spectrometry.
440 *Proteomics*. 2008;8:3715-24.

441 [31] Lemaire R, Desmons A, Tabet JC, Day R, Salzet M, Fournier I. Direct analysis and MALDI imaging of
442 formalin-fixed, paraffin-embedded tissue sections. *J Proteome Res*. 2007;6:1295-305.

443 [32] Nilsson A, Fehniger TE, Gustavsson L, Andersson M, Kenne K, Marko-Varga G, et al. Fine Mapping the
444 Spatial Distribution and Concentration of Unlabeled Drugs within Tissue Micro-Compartments Using
445 Imaging Mass Spectrometry. *PLoS ONE*. 2010;5:e11411.

446 [33] Stoeckli M, Staab D, Schweitzer A. Compound and Metabolite Distribution Measured by MALDI Mass
447 Spectrometric Imaging in Whole-Body Tissue Sections. *Int J Mass Spectrom*. 2007;260:195-202.

448 [34] McDonnell LA, Remoortere Av, Zeijl Rv, Dalebout H, Bladergroen MR, André MD. Automated
449 Imaging MS: Toward High Throughput Imaging Mass Spectrometry. *J Proteomics*. 2010;73:1279-82.
450 [35] Cazares LH, Troyer D, Mendrinis S, Lance RA, Nyalwidhe JO, Beydoun HA, et al. Imaging Mass
451 Spectrometry of a Specific Fragment of Mitogen-Activated Protein Kinase/Extracellular Signal-Regulated
452 Kinase Kinase Kinase 2 Discriminates Cancer from Uninvolved Prostate Tissue. *Clin Cancer Res*.
453 2009;15:5541-51.
454 [36] Schwamborn K, Krieg RC, Reska M, Jakse G, Knuechel R, Wellmann A. Identifying Prostate Carcinoma
455 by MALDI-Imaging. *Int J Mol Med*. 2007;20:155-9.
456 [37] Chaurand P, Fouchécourt S, DaGue BB, Xu BJ, Reyzer ML, Orgebin-Crist M-C, et al. Profiling and
457 Imaging Proteins in the Mouse Epididymis by Imaging Mass Spectrometry. *Proteomics*. 2003;3:2221-223.
458 [38] Sugiura Y, Taguchi R, Setou M. Visualization of Spatiotemporal Energy Dynamics of Hippocampal
459 Neurons by Mass Spectrometry during a Kainate-Induced Seizure. *PLoS ONE*. 2011;6:e17952.
460 [39] Jones EA, van Remoortere A, van Zeijl RJM, Hogendoorn PCW, Boveé JVMG, Deelder AM, et al.
461 Multiple Statistical Analysis Techniques Corroborate Intratumor Heterogeneity in Imaging Mass
462 Spectrometry Datasets of Myxofibrosarcoma. *PLoS ONE*. 2011;6:e24913.
463 [40] Chughtai K, Heeren RMA. Mass Spectrometric Imaging for Biomedical Tissue Analysis. *Chem Rev*.
464 2010;110:3237-77.
465 [41] van Remoortere A, van Zeijl R, van den Oever N, Franck J, Longuespée R, Wisztorski M, et al. MALDI
466 Imaging and Profiling MS of Higher Mass Proteins From Tissue. *J Am Soc Mass Spectrom* 2010;21:1922-9.
467 [42] Djidja MC, Francese S, Loadman PM, Sutton CW, Scriven P, Claude E, et al. Detergent addition to
468 tryptic digests and ion mobility separation prior to MS/MS improves peptide yield and protein
469 identification for in situ proteomic investigation of frozen and formalin-fixed paraffin-embedded
470 adenocarcinoma tissue sections. *Proteomics*. 2009;9:2750-63.
471 [43] McLean JA, Ridenour WB, Caprioli RM. Profiling and imaging of tissues by imaging ion mobility-mass
472 spectrometry. *J Mass Spectrom*. 2007;42:1099-105.
473 [44] Jungmann JH, Heeren RMA. Emerging Technologies in Mass Spectrometry Imaging. *J Proteomics*.
474 2012;submitted to imaging MS special issue.
475 [45] Jones EA, Dieninger S-O, Hogendoorn PCW, Deelder AM, McDonnell LA. Imaging Mass Spectrometry
476 Statistical Analysis. *J Proteomics*. 2012.
477 [46] Balluff B, Elsner M, Kowarsch A, Rauser S, Meding S, Schumacher C, et al. Classification of HER2/neu
478 Status in Gastric Cancer Using a Breast-Cancer Derived Proteome Classifier. *J Proteom Res*. 2010;9:6317-
479 22.
480 [47] Balluff B, Rauser S, Meding S, Elsner M, Schoene C, Feuchtinger A, et al. MALDI Imaging Identifies
481 Prognostic Seven-Protein Signature of Novel Tissue Markers in Intestinal-Type Gastric Cancer. *Am J*
482 *Pathol*. 2011;179:2720-9.
483 [48] Djidja M-C, Claude E, Snel MF, Scriven P, Francese S, Carolan V, et al. MALDI-ion mobility separation-
484 mass spectrometry imaging of glucose-regulated protein 78 kDa (Grp78) in human formalin-fixed,
485 paraffin-embedded pancreatic adenocarcinoma tissue sections. *J Proteome Res*. 2009;8:4876-84.
486 [49] Lemaire R, Menguellet SA, Stauber J, Marchaudon V, Lucot JP, Collinet P, et al. Specific MALDI
487 Imaging and Profiling for Biomarker Hunting and Validation: Fragment of the 11S Proteasome Activator
488 Complex, Reg α Fragment, is a New Potential Ovary Cancer Biomarker. *J Proteome Res*. 2007;6:4127-34.
489 [50] Rauser S, Marquardt C, Balluff B, Deininger SO, Albers C, Belau E, et al. Classification of HER2
490 Receptor Status in Breast Cancer Tissues by MALDI Imaging Mass Spectrometry. *J Proteome Res*.
491 2010;9:1854-63.
492 [51] Schwartz SA, Weil RJ, Johnson MD, Toms SA, Caprioli RM. Protein Profiling in Brain Tumors Using
493 Mass Spectrometry: Feasibility of a New Technique for the Analysis of Protein Expression. *Clin Cancer*
494 *Res*. 2004;10:981-7.

495 [52] Caldwell RL, Opalenik SR, Davidson JM, Caprioli RM, Nanney LB. Tissue profiling MALDI mass
496 spectrometry reveals prominent calcium-binding proteins in the proteome of regenerative MRL mouse
497 wounds. *Wound Repair Regen.* 2008;16:442-9.

498 [53] Chaurand P, Latham JC, Lane KB, Mobley JA, Polosukhin VV, Wirth PS, et al. Imaging mass
499 spectrometry of intact proteins from alcohol-preserved tissue specimens: bypassing formalin fixation. *J*
500 *Proteome Res.* 2008;7:3543-55.

501 [54] Oppenheimer SR, Mi D, Sanders ME, Caprioli RM. Molecular Analysis of Tumor Margins by MALDI
502 Mass Spectrometry in Renal Carcinoma. *J Proteome Res.* 2010;9:2182-90.

503 [55] Taverna D, Nanney LB, Pollins AC, Sindona G, Caprioli R. Multiplexed molecular descriptors of
504 pressure ulcers defined by imaging mass spectrometry. *Wound Repair Regen.* 2011;19:734-44.

505 [56] Hardesty WM, Kelley MC, Mi D, Low RL, Caprioli RM. Protein Signatures for Survival and Recurrence
506 in Metastatic Melanoma. *J Proteomics.* 2011;74:1002-14.

507 [57] Reyzer ML, Caldwell RL, Dugger TC, Forbes JT, Ritter CA, Guix M, et al. Early Changes in Protein
508 Expression Detected by Mass Spectrometry Predict Tumor Response to Molecular Therapeutics *Cancer*
509 *Res.* 2004;64:9093–100.

510 [58] Schramm T, Hester A, Klinkert I, Both J-P, Heeren RMA, Brunelle A, et al. imzML – a Common Format
511 for Flexible Exchange and Processing of Mass Spectrometry Imaging Data. *J Proteomics.* 2012;submitted
512 to special issue.

513 [59] Goodwin RJA. Sample preparation for mass spectrometry imaging: Small mistakes can lead to big
514 consequences. *J Proteomics.* 2012;in press.

515 [60] Bell AW, Deutsch EW, Au CE, Kearney RE, Beavis R, Sechi S, et al. A HUPO test sample study reveals
516 common problems in mass spectrometry–based proteomics. *Nat Methods.* 2009;6:423-30.

517 [61] Friedman DB, Andacht TM, Bunger MK, Chien AS, Hawke DH, Krijgsveld J, et al. The ABRF Proteomics
518 Research Group Studies: Educational exercises for qualitative and quantitative proteomic analyses.
519 *Proteomics.* 2011;11:1371-81.

520 [62] Martinez-Bartolome S, Blanco F, Albar J-P. Relevance of proteomics standards for the ProteoRed
521 Spanish organization. *J Proteomics.* 2010;73:1061-6.

522 [63] Végvári Á, Rezeli M, Sihlbom C, Häkkinen J, Carlsohn E, Malm J, et al. Molecular microheterogeneity
523 of prostate specific antigen in seminal fluid by mass spectrometry. *Clin Biochem.* 2012;45:331-8.

524 [64] Altelaar AFM, van Minnen J, Jiménez CR, Heeren RMA, Piersma SR. Direct Molecular Imaging of
525 *Lymnaea stagnalis* Nervous Tissue at Subcellular Spatial Resolution by Mass Spectrometry. *Anal Chem.*
526 2005;77:735-41.

527 [65] Burnum KE, Cornett DS, Puolitaival SM, Milne SB, Myers DS, Tranguch S, et al. Spatial and temporal
528 alterations of phospholipids determined by mass spectrometry during mouse embryo implantation. *J*
529 *Lipid Res.* 2009;50:2290-8.

530 [66] Kertesz V, Van Berkel GJ. Fully Automated Liquid Extraction-Based Surface Sampling and Ionization
531 Using a Chip-Based Robotic Nanoelectrospray Platform. *J Mass Spectrom.* 2010;45:252-60.

532

533